

## Blood Metabolic and Hematology Parameters and Survivorship in Mice after Application of the Rabies Challenge Virus Standard in Vaccine Potency Test

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### ABSTRACT

**Background:** Rabies virus can cause intensive and lethal infection of the central nervous system (CNS) in animals and humans. Metabolic examinations are conducted at the cerebrospinal fluid (CSF), and it has been found that many metabolic changes occur during RABV infection. However, although it is a neurotropic virus, it can cause damage to extraneural tissues - lungs, heart, kidneys and liver. This study aimed to determine differences in metabolic, endocrinology and hematologic parameters in blood of mice after application of rabies challenge with virus standard 27 strain (CVS-27).

**Materials, Methods & Results:** This study included 30 survived, and 30 dead mice that were part of the standard procedure of NIH (National Institute of Health) test in Pasteur Institute in Novi Sad. Tests were performed in the following order: two groups of mice were vaccinated in a 7 day period with different dilutions of standard vaccine and the examined vaccine. Seven days after the last vaccination, immunized animals and animals in the control group received test virus CVS-27. Blood samples were collected from a heart puncture. Differences in hematologic and biochemical parameters were determined by t-test. Due to a high number of blood parameters, we performed a joint analysis of multiple dependent variables. Higher pH value and higher concentrations of glucose, cholesterol, lactate dehydrogenase (LDH), creatine kinase (CK), albumin, urea, creatinine,  $\alpha$ -amylase, magnesium (Mg), nonesterified fatty acids (NEFA), beta-hydroxybutyrate (BHB) and lactate were noted in dead mice. Higher granulocytes and mean platelet volume (MPV) were noted in mice which died, but also reduced lymphocytes, erythrocytes, haemoglobin, hematocrit and platelets count. Higher values of insulin, cortisol and HOMA-IR (homeostatic model assessment insulin resistance) were noted in the group of dead mice compared to the surviving one. Reduced QUICKI (quantitative insulin sensitivity check index) value was noted in mice which died compared to the surviving group. Principal component analysis (PCA) showed that components 1 and 2 explain 38.7 of variance and that these two compounds are enough for the distinction between the animals which dies and those that survived. It was found that the cortisol, insulin, HOMAIR, NEFA, aspartate aminotransferase (AST), lactic acid, LDH and granulocyte could explain the variance of the first component, which highly correlated with the first principal component. Also, pH level, glucose, creatinine, albumin and BHB showed significant importance. A positive correlation was shown between those parameters.

**Discussion:** Mice that died during NIH test after applying CVS-27 expressed more significant stress (higher cortisol level). Disturbances of energy metabolism were noted (more significant catabolism of lipids and insulin resistance), changes of protein metabolism caused by muscle load (urea, creatinine, AST and LDH) and general disturbances of acid-base status (higher pH) and dehydration (increased albumin) were also noted in mice that died. Values of hematologic parameters showed minor influence at total variability and are a bit correlated with metabolic changes. In factor analysis, component 1 was determined from numerous parameters. Correlations between component 1 and cortisol, HOMA-IR, lactates, insulin, AST and LDH were noted. It completely determines survived and dead animals after CVS-27 during NIH test. Disturbances in blood parameters showed an analogy with previous studies of CNS. Given parameters can be very useful in clinical-pathological analysis in RABV infection.

**Keywords:** mice, rabies virus, blood metabolite.

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## INTRODUCTION

Rabies virus (Family of *Rhabdoviridae*, Genus *Lyssavirus*) causes highly intensive and lethal infections in people and animals. It is a neurotropic virus with a high affinity for cell receptors, cytoplasm and transports across interneuronal synapses [8]. Rabies virus causes fatal encephalitis followed by a response of other organ systems. RABV stimulates immunologic response [3, 22], but there are no great pathologic changes in CNS and immunological clearance of RABV. The main cause of the lethal effect is the inhibition of the synthesis of proteins that showed a crucial role in cell metabolic functions [9]. Previous results showed significant metabolic changes in cerebrospinal fluid (CSF) during viral infection [32–34]. Metabolomic methods of CSF showed the occurrence of ketosis, while in later stages, there were changes in protein metabolism and cytotoxicity [21]. Differences in gene expression and domination of anaerobic metabolism in CNS during RABV infection were confirmed [23]. Even though the Rabies virus is neurotropic, some research showed that it could cause changes in many other tissues - lungs, heart, liver and kidneys [16]. Those results showed that macrophages are very significant in extraneural viral dissemination. Also, systemic response to RABV infection includes greater blood corticosterone concentrations [26] or systemic metabolic adaptations like keto-alkalosis to be found [7].

This study aims to determine differences in the systemic response of metabolic, endocrine and hematologic parameters in blood of mice that survived and not survived the standard National Institutes of Health (NIH) test [15] and the application of challenge virus standard 27 strain (CVS-27).

## MATERIALS AND METHODS

### *Animals and treatment*

The study included 30 mice that survived and 30 that did not. The study was conducted at Pasteur Institute in Novi Sad. The level of protection that is induced by the inactivated vaccine was determined by test. NIH test was performed on 2 mice groups at an interval of 7 days, with different standard and vaccine solutions. Seven days after the last vaccination, animals from the immunized and the control group received a test virus (Challenge Virus Standard, strain CVS-27)<sup>1</sup>. Mice<sup>2</sup> (white mice, strain Naval Medical Research Institute-NMRI) were supervised in the period of 14 days and mean effective

dose (ED50) was determined according to the number of surviving animals. Before the test, animals were selected so that all animals included in the experiment were at least 4-week-old, 23-27 g weight and in good health condition. Before intracerebral inoculation of CVS-27 virus, in order to perform NIH test, animals were anaesthetized and treated with antibiotics administered intraperitoneally (the anaesthetic solution contained 1% ketamine and 0.1% xylazine diluted with sterile 0.01 mol/L PBS and antibiotics).<sup>3</sup> The amount of anaesthetic dilution per mouse was 0.3 mL. The anaesthetic solution included 0.1 mL per 10g of body weight. After 3 min, every mouse received 0.05 mL/ 10 g BW of anaesthetic solution, and this was repeated until animals were fully anaesthetized. During full anaesthesia of mice, CVS-27 was administered intracerebrally. Mice were placed in cages in lateral positions in a dark room. The complete clinical exam should be performed before inoculation. Animals who expressed difficulties in motion, ectoparasites, and changes in fur or diarrhoea were excluded from the study.

### *Blood parameters*

Blood samples were collected by heart puncture in appropriate tubes.<sup>4</sup> Individual blood samples were taken after terminal anaesthesia on animals that survived at the end of the experiment. pH level was determined by pH-meter.<sup>5</sup> Hematologic parameters were determined by automatic analyzer<sup>6</sup> using software for lab mice. Biochemical parameters were measured by standard reagents<sup>7</sup> at automatic spectrophotometer<sup>8</sup>. Insulin and cortisol concentration were measured by automatic endocrinology analyzer<sup>9</sup>. Insulin resistance indexes QUICKI, HOMA- $\beta$  and HOMA-IR were determined according to formulas for lab mice [2].

### *Statistical analysis*

Differences in values of hematologic and biochemical parameters were determined by *t*-test. Due to a high number of blood parameters, we performed a joint analysis of multiple dependent variables. Different metabolite levels in mice that survived and died after rabies virus challenge were illustrated by heatmap. The development of class model based on blood parameters as a predictor of survivorship in mice after rabies virus challenge in NIH test was performed by principal component analysis (PCA) with score plot and loading plot presentation. Statistical software<sup>10</sup> SPSS (USA) and online platform<sup>11</sup> were used for analysis and results representation.

RESULTS

Significant differences in values of haematological, biochemical and endocrine parameters were determined in mice that died after RCVS application during NIH test compared to the control group (Table 1). Higher levels of glucose, cholesterol, LDH, CK, albumin, urea, creatinine, lactate, amylase, Mg, NEFA, BHB and higher pH value were determined in dead mice. Greater neutrophil count and MPV, and reduced lymphocytes, erythrocytes, haemoglobin, hematocrit and platelets were found in dead animals. Endocrine changes in dead animals compared to control were: higher insulin, cortisol and HOMA-IR index, while QUICKI was reduced. No significant deviation of other parameters was noted.

The magnitude of the change in lab parameters is shown at the heat map (Figure 1). The heat map of the changes in metabolites related to survivorship of mice - blue colour represents the trend of decrease, and red represents a rising trend. The heat map shows differences in expression of numerous parameters in mice that survived and died, but some of the parameters do not express this when considered for each animal itself (MCV, P, Mg, Ca, WBC, TBil, PLT).

Principal component analysis (PCA) shows that component 1 and 2 explain 38.7% of the variance and that these two components are enough for proper discrimination of mice that died and survived (Figure 2). It shows that survivorship could be classified by PCA scores, and discrimination is mainly determined by the X-axis (PC1).

As shown in Figure 3, the two axes represent the first 2 components, namely component 1 and component 2. It was found that cortisol, insulin, HOMA-IR, NEFA, AST, lactic acid, LDH and granulocyte could explain the variance of the first component, and highly correlated with the first principal component (major distance from the intersection point of two lines that pass 0 at X and Y-axis). Minor compared to this, but still, significant influence has pH, glucose, creatinine, albumin and BHB (minor distance from the intersection point). These parameters show a positive correlation that can be reflected in a sharp angle whose top is the intersection point of lines that pass thru 0 and arms pass dots of examined parameters.

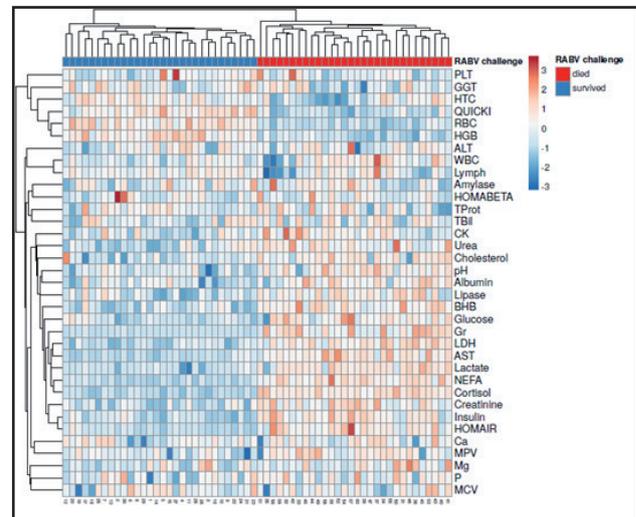


Figure 1. Heat map of expression of blood parameters in dead and surviving mice after RABV challenge (application of CSV-27).

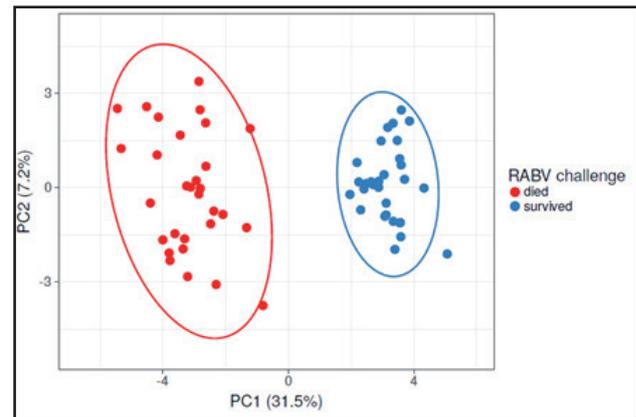


Figure 2. Principal component analysis and classification of died and survived mice after RABV challenge (application of CSV-27) in accordance to principal component 1 and 2 (PC1 and PC2).

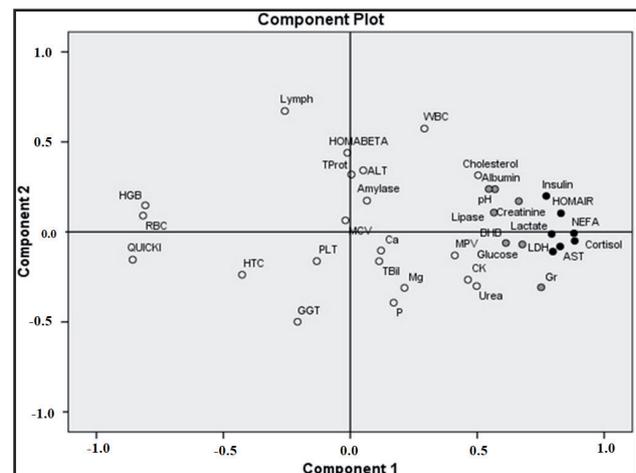


Figure 3. Biplot of component coefficients for each blood variable.

**Table 1.** Blood parameters in died and survived mice after application of rabies virus standard challenge (CVS-27).

Blood parameter	Survived		Died		Significance
	Mean	SD	Mean	SD	
pH	7.36	0.08	7.47	0.08	$P < 0.01$
Glucose (mmol/L)	8.82	1.38	11.30	2.27	$P < 0.01$
Cholesterol (mmol/L)	2.69	0.27	2.99	0.27	$P < 0.01$
LDH (IU/L)	542.50	87.90	782.59	108.31	$P < 0.01$
AST (IU/L)	89.23	15.74	163.33	34.49	$P < 0.01$
CK (IU/L)	89.23	15.76	106.19	18.76	$P < 0.01$
Urea (mmol/L)	7.48	0.66	8.30	0.67	$P < 0.01$
Creatinine ( $\mu\text{mol/L}$ )	27.15	2.54	31.50	2.94	$P < 0.01$
Lactate (mmol/L)	4.50	0.39	5.49	0.48	$P < 0.01$
NEFA (mmol/L)	0.13	0.04	0.35	0.08	$P < 0.01$
BHB (mmol/L)	0.28	0.10	0.45	0.10	$P < 0.01$
Albumin (g/L)	26.40	2.84	29.66	2.66	$P < 0.01$
TProt (g/L)	47.91	3.92	47.91	5.55	NS
Ca (mmol/L)	2.51	0.20	2.54	0.21	NS
P (mmol/L)	3.09	0.25	3.20	0.22	NS
Mg (mmol/L)	0.55	0.06	0.58	0.06	$P < 0.05$
ALT (IU/L)	35.27	3.50	35.70	3.89	NS
GGT (IU/L)	5.28	0.45	5.07	0.51	NS
TBil ( $\mu\text{mol/L}$ )	3.89	0.68	4.04	0.71	NS
Amylase (IU/L)	917.33	99.19	922.07	103.96	NS
Lipase (IU/L)	83.39	9.02	93.90	8.34	$P < 0.01$
WBC ( $\times 10^9/\text{L}$ )	15.45	1.10	16.25	2.12	NS
Gr ( $\times 10^9/\text{L}$ )	2.10	0.18	4.18	1.29	$P < 0.01$
Lymph ( $\times 10^9/\text{L}$ )	13.35	1.04	12.07	2.67	$P < 0.05$
RBC ( $\times 10^{12}/\text{L}$ )	12.41	1.47	8.81	1.04	$P < 0.01$
Hemoglobin (g/L)	165.37	7.97	145.52	7.02	$P < 0.01$
MPV (fL)	4.71	0.57	5.24	0.64	$P < 0.01$
Hematocrit HTC (%)	43.55	1.35	41.98	2.27	$P < 0.01$
MCV (fL)	45.08	0.97	45.08	0.97	NS
PLT ( $\times 10^9/\text{L}$ )	764.17	70.86	758.10	68.37	NS
Cortisol (ng/mL)	7.68	1.52	16.47	2.77	$P < 0.01$
Insulin (ng/mL)	0.53	0.13	0.84	0.17	$P < 0.01$
HOMAbeta	2.18	0.98	2.19	0.75	NS
HOMAIR	0.21	0.06	0.42	0.14	$P < 0.01$
QUICKI	0.53	0.03	0.45	0.03	$P < 0.01$

## DISCUSSION

NIH test is a referent test for the evaluation of the potency of the anti-rabies vaccine. This test includes live lab mice, whose blood serves as a biologic

material to examine differences in blood parameters disturbances in surviving and dead mice. NIH test is a standard test which has been used since the '70s [27]. Due to welfare 3R rules, there are some alternatives

to this test [29]. Most of the vaccines are made of strains Pasteur virus (PV), Pitman-Moore (PM) and challenge virus standard (CVS). In studies of vaccine potency, CVS strain was compared with other strains of rabies virus significant in veterinary medicine. Some specifics of CVS usage have been found in the process of evaluation of vaccine potency [28]. Changes in CNS show that apoptosis is significant in the process of cell death. The more the virus is neurovirulent, the apoptosis is less expressed [18]. One of the ways of cell degeneration is excitotoxicity. This is the pathological process by which neurons are damaged and killed by the over-activation of receptors. So the brain damage in rabies is based on metabolic changes. Some studies have shown that viral RNA could be found in blood samples of infected mice. This indicates that viremia could develop during rabies infection [17].

Cases of transmission from man to man during transplantation of organs and cornea are described in the literature [19]. All of the listed could be related to the systemic presence of rabies virus in an organism. This could explain metabolic adaptations in mice that survived and did not survive NIH test.

A higher concentration of cortisol, insulin and disturbance in insulin sensitivity were found in mice that died after the application of CVS-27 during NIH test. That indicates stress response and insulin resistance development. Studies are mainly based on corticosterone concentrations, but newer studies show a high correlation of cortisol and corticosterone in lab mice during physiologic, acute, and chronic stress [11]. Cortisol has numerous roles in the organism. Cortisol increases the level of gluconeogenesis [4], which can be related to increased glycaemia in lab mice that died after the application of CVS-27. Earlier works showed that in the early phase of rabies, glucosuria, and hyperglycaemia could be noticed [20]. Higher glycaemia follows an increase in insulin, but it was noted that there was insulin resistance development in mice that died. In lab mice, acute stress or cortisol application can cause an increase in insulin level [1]. It has been shown that Brain-to-Pancreatic Islet Neuronal Map allows CNS to cause the increased secretory function of pancreas. That is proven by the usage of pseudorabies virus [24]. These regulations can be related to increased insulin concentration in the blood of mice that died after CVS-27 inoculation. Insulin resistance is defined as the incapability of pancreatic cells to produce insulin

or inadequate tissue response to circulating insulin [6]. Insulin resistance in lab mice is related to higher HOMA-IR index and lower QUICKI index [2]. Our results are in accordance with theirs. Insulin resistance is followed by greater lipolysis and ketogenesis because insulin is the only lipogenic hormone.

Other hormones, including cortisol, are lipolytic. During the NIH test in later stages, mice expressed reduced food intake and starvation that caused increased lipolysis and a decrease in glucose and insulin [14]. Higher concentrations of glucose and insulin were noted in dead mice in our research. A higher concentration of glucose and ketone bodies in CNS is found in later CNS infection stages [26]. Obtained results are in accordance with this. Same authors concluded that during RABV infection, CNS uses less glucose and insulin for metabolic processes. Changes that were noted in our study are similar to changes shown in CNS during RABV infection.

The large group of metabolites showed a smaller but still significant effect in explaining metabolic variance after CVS-27 application. The increase in pH of dead animals was noted, which is in accordance with keto-alkalosis development in one described case of rabies in people [7]. Alkalosis in rabies is of a respiratory type, and it is developed by hyperventilation, inspiratory spasm and deeper inspiration. That can be related to encephalitis, followed by an increase in alveolar pressure and development of pneumomediastinum [12,31]. Increased concentrations of ALT and LDH could be caused by stress [25] and more significant muscle load. RT-PCR and immunohistochemistry methods showed that wild strain of rabies virus could replicate in muscular tissue [5]. Muscle load can be related to neuromuscular irritability and spasm. RABV is replicated in muscles, so direct action of viral particles cannot be excluded [13]. Higher values of urea and creatinine could be related to protein catabolism and muscle load. Higher albumin values could be related to dehydration of the organism and great loss of water. Diabetes insipidus is developed in encephalitis syndrome [30], and it is characterized by greater water loss with decreased kidney filtration. Lactate concentrations were increased in dead mice. CSF examinations show higher lactate concentrations in mice infected with RABV, especially in later stages [21]. Higher lactate concentration could be a consequence of changes in the blood-brain barrier that develops during rabies.

Haematological changes in rabies are mild. In our research, they are greater and can be related to cortisol and stress action [4]. MPV change can be a sign of immunologic thrombocytopenic purpura described after rabies vaccine application [10].

#### CONCLUSIONS

Mice that died during NIH test after applying CVS-27 expressed more significant stress (higher cortisol level). Disturbances of energy metabolism were noted (more significant catabolism of lipids and insulin resistance), changes of protein metabolism caused by muscle load (urea, creatinine, AST and LDH) and general disturbances of acid-base status (higher pH) and dehydration (increased albumin) were also noted in mice that died.

This study demonstrated that PCA was capable of differentiating between died and survived mice by measuring their blood biochemical parameters. Discriminant function analysis was able to clearly identify died mice after application of CVS-27 using the biochemical parameters (cortisol, HOMA-IR, lactates, insulin, AST and LDH) as input data which strongly correlates with principal component 1.

Hematology parameters showed mild effect on metabolic parameters variability and survival

outcome in mice. Disturbances in blood metabolic parameters showed an analogy with previous studies of CNS. Blood parameters can be very useful in clinical-pathological analysis in RABV infection.

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**Ethical approval.** Experiment was conducted in accordance with the decision and authorization of the Ministry of Agriculture, Forestry and Water Management (Veterinary Administration) number 323-07-01465/2014-05-1.

**Declaration of interest.** The authors report no conflicts of interest. The authors alone are responsible for the content and writing of paper.

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